



PDF hosted at the Radboud Repository of the Radboud University Nijmegen

The following full text is a publisher's version.

For additional information about this publication click this link.

<http://hdl.handle.net/2066/191177>

Please be advised that this information was generated on 2018-06-17 and may be subject to change.

Modular, Bioorthogonal Strategy for the Controlled Loading of Cargo into a Protein Nanocage

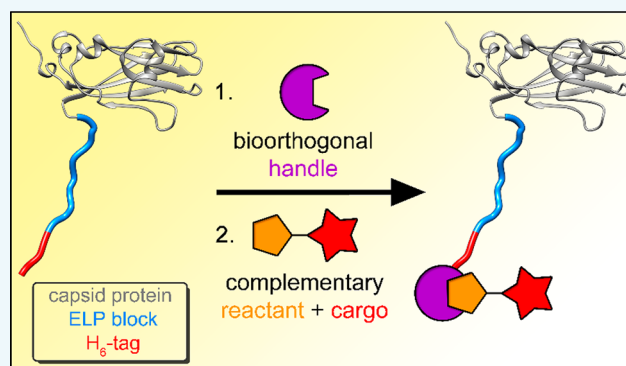
Lise Schoonen,^{†,‡,§} Selma Eising,^{§,¶} Mark B. van Eldijk,[†] Jaleesa Bresseleers,[†] Margo van der Pijl,[†] Roeland J. M. Nolte,[†] Kimberly M. Bongers,^{*,§,¶} and Jan C. M. van Hest^{*,†,‡,§,¶}

[†]Department of Bio-Organic Chemistry and [§]Department of Biomolecular Chemistry, Institute for Molecules and Materials, Radboud University, Heyendaalseweg 135, 6525 AJ Nijmegen, The Netherlands

[‡]Laboratory of Bio-Organic Chemistry, Eindhoven University of Technology, PO Box 513 (STO 3.31), 5600 MB Eindhoven, The Netherlands

Supporting Information

ABSTRACT: Virus capsids, i.e., viruses devoid of their genetic material, are suitable nanocarriers for biomedical applications such as drug delivery and diagnostic imaging. For this purpose, the reliable encapsulation of cargo in such a protein nanocage is crucial, which can be accomplished by the covalent attachment of the compounds of interest to the protein domains positioned at the interior of the cage. This approach is particularly valid for the capsid proteins of the cowpea chlorotic mottle virus (CCMV), which have their N-termini located at the inside of the capsid structure. Here, we examined several site-selective modification methods for covalent attachment and encapsulation of cargo at the N-terminus of the CCMV protein. Initially, we explored approaches to introduce an N-terminal azide functionality, which would allow the subsequent bioorthogonal modification with a strained alkyne to attach the desired cargo. As these methods showed compatibility issues with the CCMV capsid proteins, a strategy based on 2-pyridinecarboxaldehydes for site-specific N-terminal protein modification was employed. This method allowed the successful modification of the proteins, and was applied for the introduction of a bioorthogonal vinylboronic acid moiety. In a subsequent reaction, the proteins could be modified further with a fluorophore using the tetrazine ligation. The application of capsid assembly conditions on the functionalized proteins led to successful particle formation, showing the potential of this covalent encapsulation strategy.



INTRODUCTION

From a nanotechnological perspective, viruses have been recognized as interesting carrier tools, due to their unique ability to protect their nucleic acid cargo and deliver it to cells in their infected host. In particular, virus-like particles (VLPs), viruses that are devoid of their viral nucleic acids, are used for nanocarrier purposes, as they are relatively harmless without their viral genetic information and have room in their interior for loading of cargo such as catalysts, drugs, or imaging agents.

A remarkable example of a VLP that has proven very suitable as a nanocarrier is the cowpea chlorotic mottle virus (CCMV) capsid. In contrast to most other VLPs, CCMV capsids show reversible assembly and disassembly behavior, even in the absence of their genetic material. The capsid proteins exist as dimers in solution at a physiological pH and spontaneously form 28-nm-sized capsids with $T = 3$ symmetry out of 90 capsid protein dimers when the pH is lowered to pH 5.0.^{1,2} In order to make the capsids more stable at neutral pH for *in vivo* applications, we previously introduced an elastin-like polypeptide (ELP) block at the N-terminus of the CCMV capsid protein.³ ELPs are stimulus-responsive polypeptides, consisting

of repeating Val-Pro-Gly-Xaa-Gly (VPGXG) pentapeptides containing any natural amino acid guest residue, Xaa (X), except proline.^{4,5} These peptides can reversibly switch from a water-soluble state to a collapsed, hydrophobic state upon a change in the environmental conditions. The introduction of this peptide at the N-terminus of the CCMV capsid protein resulted in a new ELP-induced assembly pathway that is triggered by increasing the temperature or salt concentration, yielding smaller $T = 1$ capsids with a diameter of 18 nm, composed of 30 capsid protein dimers.³ The endogenous assembly pathway leading to $T = 3$ capsids could still be triggered by lowering the pH to 5.0. The development of this ELP-CCMV variant resulted in a wider range of conditions at which the capsids remained stable, showing promise for *in vivo* applications.

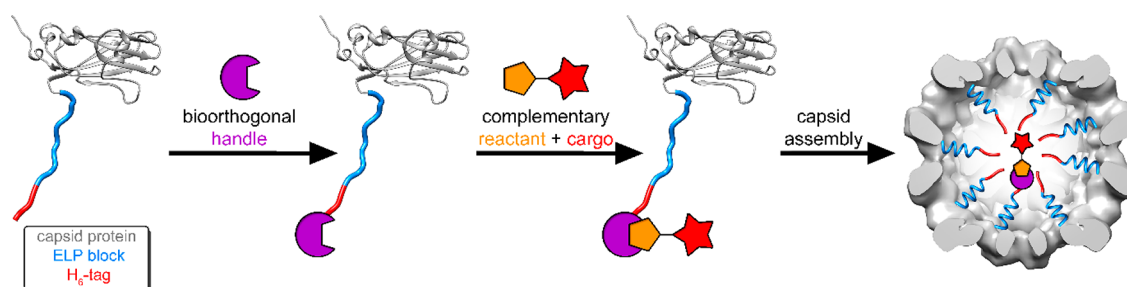
Controlled encapsulation of cargo in the ELP-CCMV capsid interior is crucial for its successful application as a nanocarrier.

Received: December 21, 2017

Revised: January 31, 2018

Published: February 6, 2018

Scheme 1. Schematic Representation of the Two-Step Modular Protocol for N-Terminal Modification of ELP-CCMV Capsid Proteins^a



^aFirst, a bioorthogonal handle is site-specifically introduced at the N-terminus. Subsequently, addition of a complementary handle modified with the desired cargo, results in covalent attachment of the cargo to the capsid proteins. Upon capsid assembly, the cargo is then encapsulated in the interior of the resulting ELP-CCMV capsids.

A classical way to encapsulate cargo in CCMV capsids is via statistical encapsulation, where cargo is added to the capsid proteins when they are in the dimer state, after which assembly conditions are applied to induce capsid formation. This, however, normally yields partial encapsulation, while most of the cargo material is lost.⁶ To increase the encapsulation efficiency, the cargo can be equipped with negatively charged moieties, mimicking the charge of the endogenous nucleic acid cargo and allowing interactions with the cationic N-terminal tails of the CCMV capsid proteins.^{7–9} The latter method, however, is not applicable for cargo encapsulation in ELP-CCMV capsids, as the ELP sequence was inserted at the N-terminus in such a way that it replaced the cationic RNA-binding motif. Alternative methods that have been used for cargo encapsulation in CCMV VLPs are (i) genetic cloning of a target protein onto the N-terminus of the capsid proteins;¹⁰ (ii) the genetic introduction of an N-terminal domain, which can form noncovalent interactions with a specific complementary domain;^{11,12} and (iii) enzymatic modification of the N-terminus using the enzyme Sortase A.^{13,14} The latter methodology is possibly the most optimal for versatile and reproducible cargo encapsulation, since it is modular, does not require extensive genetic engineering, and yields a robust covalent link between the cargo and the capsid proteins. Even though this technique is applicable to any cargo that can be equipped with the required Sortase recognition sequence, it lends itself best to the encapsulation of peptides and proteins, as these allow easy introduction of this recognition peptide. In order to further broaden the modularity and possibilities for cargo loading, we set out to develop a method for the selective modification of the ELP-CCMV capsid protein N-termini, which eliminates the need for incorporation of a peptide into the cargo and is therefore more easily applied to small molecules such as (organo)catalysts or imaging agents.

In order to achieve site-specific N-terminal modification, we aimed to introduce a reactive handle onto the N-terminus that can be used in a bioorthogonal reaction, a reaction that is inert to any natural occurring biological functionalities, such as the copper-catalyzed alkyne–azide cycloaddition (CuAAC), strain-promoted alkyne–azide cycloaddition (SPAAC), or the inverse electron-demand Diels–Alder (iEDDA) reaction with tetrazines.^{15–17} Fortunately, several methodologies are available to achieve the site-selective modification of the N-terminal amine over amines present in lysine side chains, ranging from pH-controlled reactions to reactions requiring side chain participation.¹⁸ Here, we describe our efforts to site-selectively

modify the N-terminus of the CCMV capsid protein with a bioorthogonal reagent, and subsequently attach a model cargo in a modular fashion (Scheme 1).

RESULTS AND DISCUSSION

Introduction of an Azide Function at the Capsid Protein N-Terminus. We started exploring a route toward the selective N-terminal modification of the ELP-CCMV capsid proteins by introducing an azide functionality, which is widely used as a bioorthogonal handle. Azides have been introduced into proteins cotranslationally using a genetic engineering approach,^{19–22} or post-translationally via chemical or enzymatic modification.^{23–28} Initially, we attempted the post-translational chemical modification by applying a selective diazotransfer reaction to convert the α -amine at the N-terminus into an azide.²⁴ In proteins, this primary amine often has the lowest pK_a and can therefore be modified selectively using the right conditions. The reaction of ELP-CCMV with imidazole-1-sulfonyl azide **1** was performed in a diethanolamine buffer of pH 8.5 (Figure 1A). Analysis by mass spectrometry of the N-terminal fragments, obtained by tryptic digestion, revealed a mass shift of 26 Da, corresponding to a successful diazotransfer reaction (Figure 1B). The other lysine-containing fragments were also detected and did not show a mass shift. Unfortunately, we observed partial precipitation of the CCMV protein during the reaction and a significant amount of residual starting material, even after further optimization of the reaction conditions.

The observed precipitation of CCMV in the diazotransfer reaction prompted us to explore other options for the introduction of an N-terminal azide. For this, we investigated the residue-specific biosynthetic incorporation of an unnatural amino acid by using azidohomoalanine (Aha) as a methionine surrogate.¹⁹ ELP-CCMV contains one additional methionine residue, Met137, positioned in the capsid protein part of the fusion protein. To prevent undesired incorporation of Aha at this position, we mutated Met137 into an alanine residue. In addition, we inserted an additional arginine residue following the N-terminal methionine to prevent processing of the N-terminal residue by bacterial aminopeptidases (SI - experimental section).²⁹ The new construct was first expressed in *E. coli* in the presence of methionine to confirm that the adjustments did not alter the properties of the capsid proteins and that the N-terminal methionine would indeed stay intact. Analysis of the affinity-purified Met-ELP-CCMV by SDS-PAGE and ESI-TOF mass spectrometry showed an acceptable pure

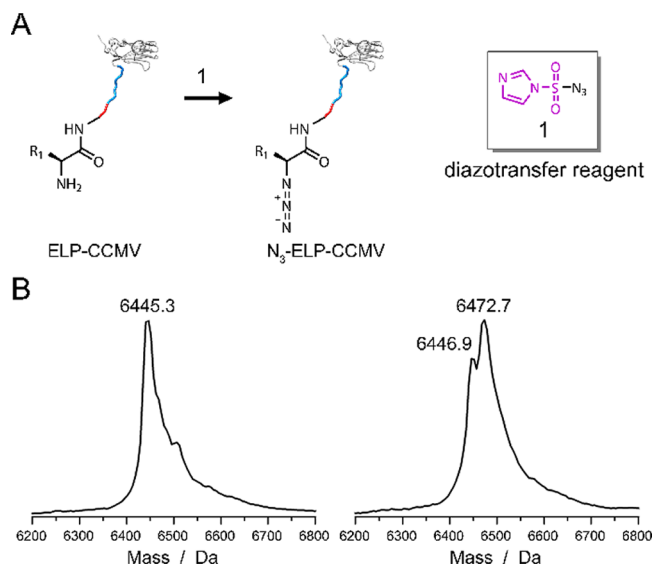


Figure 1. (A) Schematic representation of N-terminal modification of ELP-CCMV with imidazole-1-sulfonyl azide **1** to obtain N₃-ELP-CCMV. (B) MALDI-TOF mass spectra of the N-terminal tryptic fragment before (top) and after (bottom) diazotransfer. A mass shift of +26 Da is observed, corresponding to the diazotransfer of the N-terminal amine.

protein sample (Figure 2A) with the expected molecular weight (Figure S1). Investigation of the endogenous assembly pathway (lowering the pH to 5 for $T = 3$ particles) of this new protein by size exclusion chromatography coupled to multiangle laser light scattering (SEC-MALLS) showed that the VLPs had a molecular weight of 4.0 ± 0.1 MDa (Figure 2B), which is in good agreement with the $T = 3$ particles containing 180 capsid proteins of 22 570.7 Da. Further analysis with transmission electron microscopy (TEM) showed monodisperse spherical particles with a diameter of 28 nm ($T = 3$ particles), confirming that the introduced modifications did not affect the assembling properties of the ELP-CCMV (Figure 2C).

Next, we expressed the modified ELP-CCMV construct in the presence of Aha in a methionine auxotrophic *E. coli* strain. Following similar purification and analysis procedures as above

we confirmed the efficient incorporation of N-terminal Aha, indicated by the 5 Da observed mass difference upon replacement of methionine with Aha (Figure S1). Unfortunately, the protein was expressed with a 10× lower yield and SDS-PAGE analysis revealed many impurities in the obtained Aha-ELP-CCMV (Figure 2A). TEM analysis after pH-induced assembly clearly demonstrated the presence of 28-nm-diameter particles for Aha-ELP-CCMV (Figure 2C), whereas SEC analysis showed that CCMV proteins were only partially assembled, with the majority of the proteins being present in their dimer form (Figure 2B).³ Unfortunately, attempts to improve the expression yield and the purity of the azide-modified capsid proteins were unsuccessful.

Despite the fact that both the diazotransfer reaction and the genetic modification of CCMV did not yield flawless formation of azide-functionalized ELP-CCMV, we subsequently attempted to react the modified proteins further in a cycloaddition reaction. As we observed partial aggregation of the protein upon the addition of copper, which is needed for the CuAAC reaction, we decided to use the copper-free SPAAC reaction for the subsequent modification step.³⁰ To this end, both azide-modified proteins were reacted with a commercially available fluorescently labeled cyclooctyne (BCN-lissamine-rhodamine B), after which fluorescently imaged SDS-PAGE analysis showed successful modification of the N-terminus of ELP-CCMV (Figure S2). Unfortunately, we observed reoccurring precipitation of the protein during the SPAAC reaction, which led us to investigate yet an alternative approach for the site-selective modification of the capsid proteins.

Site-Specific N-Terminal Modification Using 2-Pyridinecarboxaldehydes. Recently, a bioconjugation method for specific N-terminal protein modification was described based on 2-pyridinecarboxaldehyde (2PCA) forming an N-terminal cyclic imidazolidinone condensation product with peptides and proteins (Figure 3A).³¹ This modification proceeds in aqueous environments under mild temperature and pH and as no specific amino acid residue at the N-terminus is required, it is generally applicable to many proteins. Furthermore, many new functionalities have been introduced using this method, such as affinity tags, MRI-contrasting chelators, targeting agents, and fluorophores. As this benign method might circumvent

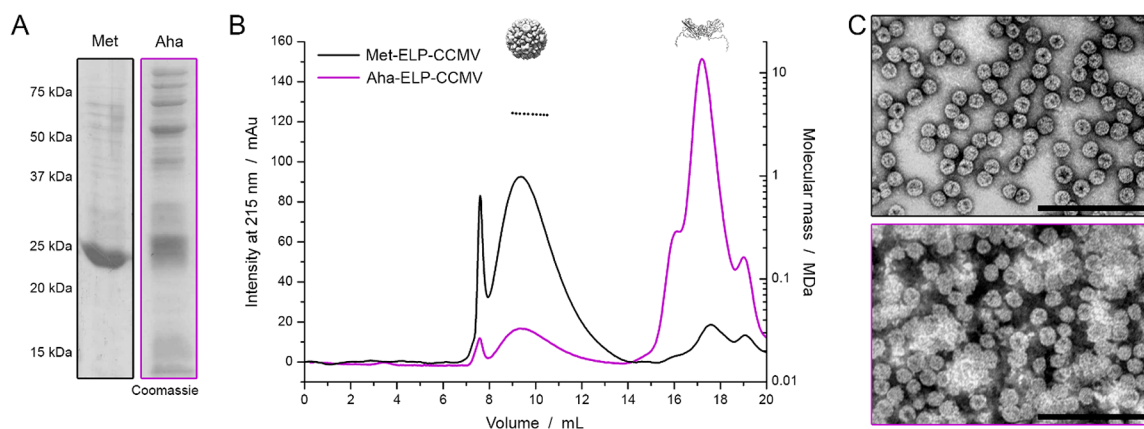


Figure 2. Characterization of Met-ELP-CCMV and Aha-ELP-CCMV. (A) SDS-PAGE analysis of Met-ELP-CCMV and Aha-ELP-CCMV after expression and Ni²⁺ affinity purification. Protein bands were visualized with Coomassie blue staining. (B) SEC-MALLS chromatograms of pH-induced assemblies of Met-ELP-CCMV (black) and Aha-ELP-CCMV (purple) measured at 215 nm. Dotted line (black) shows molecular mass data of the Met-ELP-CCMV particles. (C) Uranyl acetate-stained TEM micrographs of Met-ELP-CCMV and Aha-ELP-CCMV after pH-induced assembly. Scale bars correspond to 200 nm.

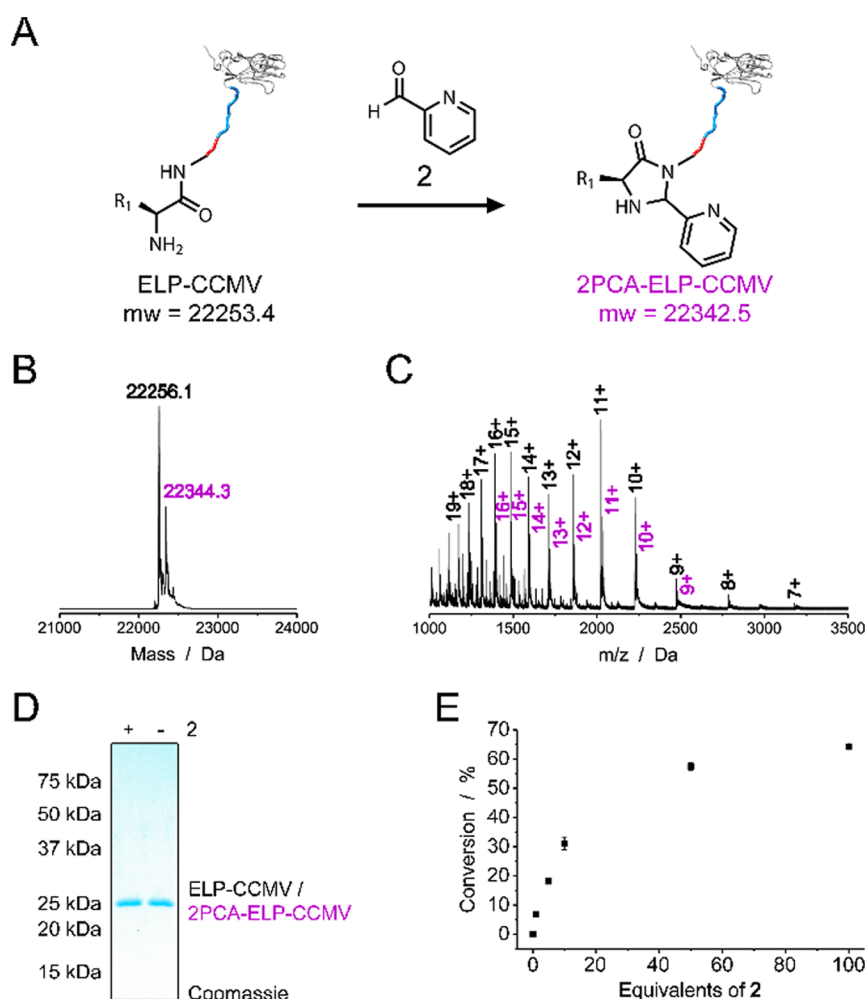


Figure 3. (A) Schematic representation of the N-terminal modification of ELP-CCMV with 2PCA 2. (B) Deconvoluted ESI-TOF mass spectrum of the modification of ELP-CCMV with 2PCA 2 (10 equiv). The expected molecular weights are 22 253.4 Da (ELP-CCMV, black) and 22 342.5 Da (2PCA-ELP-CCMV, purple). (C) Multiply charged ion series of the deconvoluted ESI-TOF mass spectrum shown in B. (D) SDS-PAGE analysis of the modification of ELP-CCMV with 2PCA 2 (10 equiv). Protein bands were stained with Coomassie blue staining. (E) Estimated conversion of ELP-CCMV to 2PCA-ELP-CCMV in reactions with 0, 1, 5, 10, 50, or 100 equiv of 2PCA 2, based on ESI-TOF analysis of the reaction mixtures. The average of duplicate measurements is plotted.

unfavorable precipitation, we aimed to evaluate this 2PCA-based modification strategy on our ELP-CCMV capsid proteins.

Initial experiments focused on the optimization of the reaction conditions for the modification of the ELP-CCMV protein with 2PCA. Following the reaction conditions as used by Francis et al. resulted in precipitation of the ELP-CCMV protein, due to the high reaction temperatures (Figure S3). This corresponded well with previous observations in our lab regarding the increased instability of the capsid proteins at elevated temperatures of 30 °C or higher. Performing the modification at room temperature, however, with 10 equiv of commercially available 2PCA 2 for 24 h in PBS buffer did result in the formation of the desired product 2PCA-ELP-CCMV, as shown by ESI-TOF analysis (Figure 3B and C). Furthermore, the modification did not result in visual precipitation of the protein, when the samples were centrifuged after the modification. SDS-PAGE analysis of the samples, of which supernatant was transferred to a clean tube before sample preparation, gave comparable intensities for both proteins, demonstrating the applicability of this modification strategy to CCMV capsid proteins (Figure 3D). To determine the highest

achievable modification yield, a series of 0, 1, 5, 10, 50, and 100 equiv of 2PCA 2 was added to the capsid proteins and the conversions were estimated using ESI-TOF analysis. Improved conversion was observed upon increasing addition of 2PCA 2 reaching a plateau of approximately 65% (Figure 3E, Figures S4, S5). Our results compare well with the research conducted by Francis and co-workers on the 2PCA modification, where conversions of 43% to >95% were achieved at 37 °C depending on the type of protein.

After having established that the modification strategy using 2PCA 2 was applicable to the ELP-CCMV capsid protein, we introduced a functional handle onto the N-terminus using this strategy. The inverse electron-demand Diels–Alder reaction of tetrazines with alkenes and alkynes is one of the most popular bioorthogonal reactions due to its selectivity and high reaction rate.^{32,33} We decided to use the reaction between vinylboronic acids (VBAs) and dipyrindyl-*s*-tetrazines, as the VBA moiety is hydrophilic, readily accessible, and stable under physiological conditions.³⁴ Additionally, it was shown previously that this bioorthogonal reaction was suitable for protein modification.

To this end, 2PCA-VBA 3 was designed, containing a 2PCA-piperazine group³¹ coupled to the pinacol protected vinyl-

boronic acid via a short linker (Figure 4A, SI - experimental section). The protected boronic ester was used instead of the

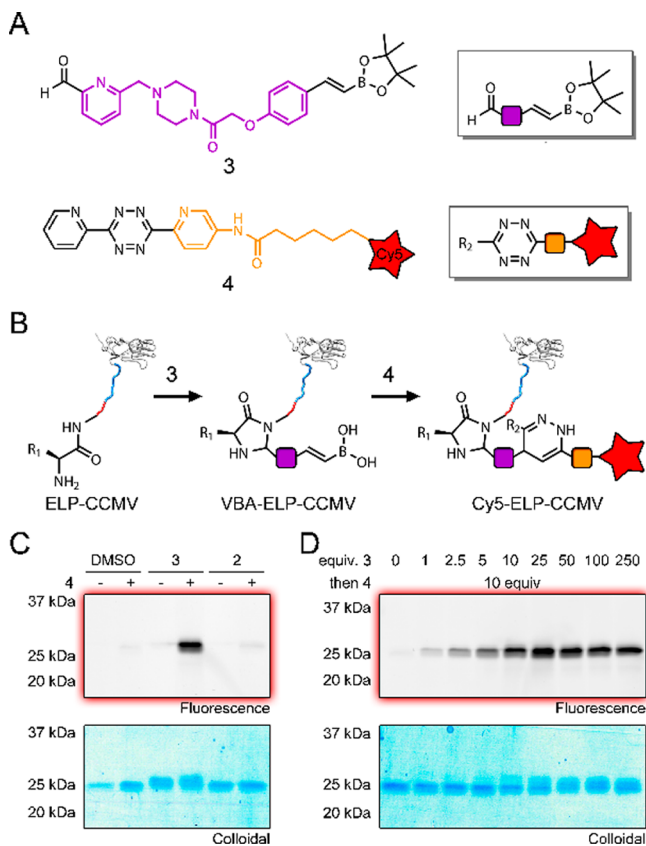


Figure 4. (A) Structures of 2PCA-VBA **3** and tetrazine-Cy5 **4**. (B) Schematic representation of N-terminal modification of ELP-CCMV with 2PCA-VBA **3**, and subsequent click reaction of tetrazine-Cy5 **4** to obtain Cy5-ELP-CCMV. (C) SDS-PAGE analysis of the modification of ELP-CCMV (10 μ M) with 2PCA **2** or 2PCA-VBA **3** (100 equiv) or DMSO as a control, and subsequent reaction with tetrazine-Cy5 **4** (10 equiv) or DMSO as a control. (D) SDS-PAGE analysis of the modification of ELP-CCMV with a concentration range of 2PCA-VBA **3** (0 to 250 equiv) and the subsequent reaction with tetrazine-Cy5 **4** (10 equiv). Protein bands were visualized by in-gel fluorescence (top) and stained with colloidal staining (bottom).

free boronic acid, since the ester is synthetically more accessible and hydrolyzes rapidly to the boronic acid in aqueous media within 15 min.³⁴ The water solubility of 2PCA-VBA **3** was found to be slightly lower than that of 2PCA **2**, and thus the concentration of ELP-CCMV capsid protein was lowered so that optimized conditions used for the coupling of 2PCA **2** could be used (100 equiv of the small molecule). The modification of ELP-CCMV using 100 equiv of 2PCA-VBA **3** was analyzed by ESI-TOF and indicated successful formation of VBA-ELP-CCMV with a yield of approximately 92% (Figure S6). Next, we performed the two-step protein modification of ELP-CCMV with 2PCA-VBA **3**, and subsequently with dipyrindyl-*s*-tetrazine **4** containing a Cy5 fluorophore as the model cargo (Figure 4A and B). The formation of Cy5-ELP-CCMV was analyzed by SDS-PAGE and showed a significant fluorescent signal for the two-step labeling using 2PCA-VBA **3** and tetrazine **4**, indicating that the coupled boronic acid is available for a subsequent reaction with a dipyrindyl-*s*-tetrazine (Figure 4C). Control reactions in which either 2PCA **2** or no 2PCA derivative (only DMSO) was used instead of 2PCA-VBA

3 showed no or only very low fluorescent signal, eliminating aspecific reactions of tetrazine-Cy5 **4** with the protein. The modification of both 2PCA-VBA **3** and tetrazine **4** caused a small mass shift of CCMV on SDS-PAGE-gel, which was too small for calculation of the modification yields. SDS-PAGE analysis of the reaction of ELP-CCMV with a concentration range of 2PCA-VBA **3** and subsequent ligation with tetrazine **4** showed that the highest achievable modification was achieved using ~50 equiv of 2PCA-VBA **3** (Figures 4D).

Finally, we investigated the assembly behavior of the modified capsid proteins. To this end, ELP-CCMV was reacted with 2PCA-VBA **3** and tetrazine-Cy5 **4**, after which the protein was washed against PBS buffer and subsequently transferred to pH 5.0 capsid buffer by centrifugal filtration, prompting pH-induced assembly of the capsid proteins. SEC analysis of the resulting solutions clearly showed a capsid peak around an elution volume of 11 mL, indicating the formation of *T* = 3 particles; no residual capsid protein dimers were observed (Figure 5). The capsid peak absorbed light of 646 nm,

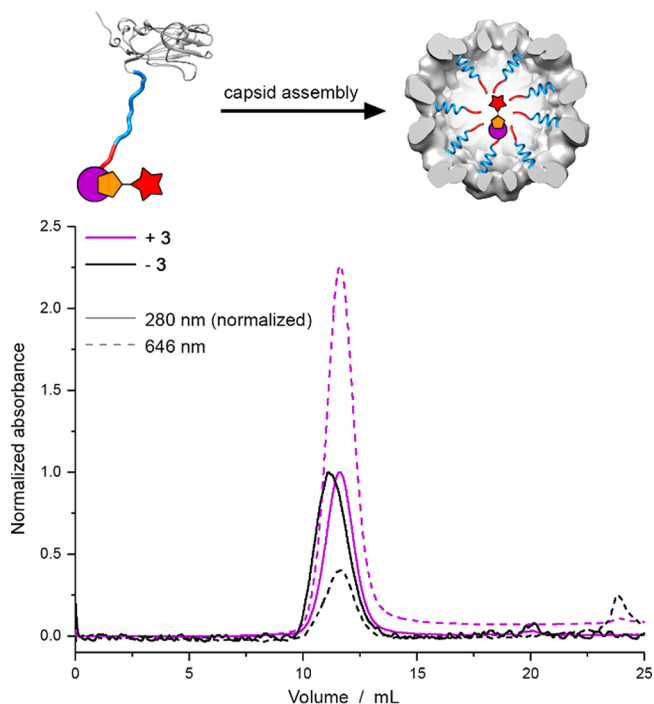


Figure 5. Size exclusion chromatogram of ELP-CCMV after modification with 2PCA-VBA **3** (purple) or DMSO as control (black). Next, both samples were treated with tetrazine-Cy5 **4**, followed by pH-induced capsid assembly. Capsids are observed around an elution volume of 11 mL. Solid lines show the absorbance of the proteins at 280 nm. Dashed lines show the absorbance of the Cy5 fluorophore at 646 nm.

demonstrating coelution with the fluorescent Cy5 dye. A control, in which 2PCA-VBA **3** had been left out of the initial modification reaction, only showed minor absorbance at 646 nm, which might result from statistical encapsulation of a small residual amount of tetrazine-Cy5 **4**.

CONCLUSIONS

N-terminal modification of the ELP-CCMV capsid proteins has proven to be challenging due to reoccurring precipitation in various reaction conditions. The diazotransfer reaction for selective modification of the N-terminal amine to an azide²⁴

resulted in significant protein precipitation, and subsequent attempts to introduce an azide-containing unnatural amino acids were not successful. Fortunately, the modification method using 2-pyridinecarboxaldehydes³¹ was found to be suitable for ELP-CCMV capsid proteins. No significant protein instability was observed during the reactions, while up to 92% of the proteins could be modified using this strategy. We applied this method to attach a bioorthogonal vinylboronic acid handle, which could be further modified with a dipyridyl-*s*-tetrazine moiety linked to a fluorescent dye as a model cargo. Proof-of-principle reactions showed that this modular two-step modification strategy was successful, demonstrating that this method is suitable for encapsulating cargo into ELP-CCMV nanocages.

■ EXPERIMENTAL PROCEDURES

Synthesis of (E)-(4-(2-(4-((6-Formylpyridin-2-yl)-methyl)piperazin-1-yl)-2-oxoethoxy)styryl)boronic acid pinacol ester (3). *tert*-Butyl 4-((6-formylpyridin-2-yl)methyl)piperazine-1-carboxylate **S7** (25 mg, 82 μ mol, 1.0 equiv) was dissolved in dry CH_2Cl_2 (1 mL) under N_2 and 4 M HCl in dioxane (205 μ L, 820 μ mol, 10.0 equiv) was added. The mixture was stirred for 2 h, whereupon the volatiles were evaporated. The solid was dissolved in DMF and (E)-(4-(2-((2,5-dioxopyrrolidin-1-yl)oxy)-2-oxoethoxy)styryl)boronic acid pinacol ester **S4** (39 mg, 98 μ mol, 1.2 equiv) was added. Then, Et_3N (34 μ L, 250 μ mol, 3.0 equiv) was added and the solution was stirred for 2 h. The volatiles were evaporated and the product was purified using column chromatography (0 to 5% MeOH in EtOAc) yielding PCA-VBA **3** (32 mg, 80%) as a white solid. R_f = 0.19 (5% MeOH in EtOAc). ^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 9.97 (s, 1H), 8.04 (t, J = 7.7 Hz, 1H), 7.84 (dd, J = 7.7, 1.1 Hz, 1H), 7.78 (dd, J = 7.7, 1.1 Hz, 1H), 7.54–7.46 (m, 2H), 7.24 (d, J = 18.4 Hz, 1H), 6.93–6.85 (m, 2H), 5.96 (d, J = 18.4 Hz, 1H), 4.85 (s, 2H), 3.75 (s, 2H), 3.51–3.44 (m, 4H), 2.53–2.49 (m, 2H), 2.45–2.40 (m, 2H), 1.23 (s, 12H). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$) δ 193.7, 165.6, 159.2, 158.9, 151.8, 148.8, 138.2, 130.0, 128.4, 127.5, 120.4, 114.8, 82.9, 65.8, 63.0, 52.8, 52.4, 44.2, 41.3, 24.7. No signal was observed for the carbon attached to boron. HRMS (ESI+) m/z calcd. for $\text{C}_{27}\text{H}_{34}\text{BN}_3\text{O}_5$ [$\text{M} + \text{H}$]⁺ 492.26698, found: 492.26703.

Synthesis of 2-((Sulfo-Cy5)amino)-N-(6-(6-(pyridin-2-yl)-1,2,4,5-tetrazin-3-yl)pyridin-3-yl)acetamide (4). 2-(Boc-amino)-N-(6-(6-(pyridin-2-yl)-1,2,4,5-tetrazin-3-yl)pyridin-3-yl)acetamide³⁴ (10.3 mg, 25 μ mol, 1.1 equiv) was dissolved in dry CH_2Cl_2 (1 mL) and 4 M HCl in dioxane (188 μ L, 0.75 mmol, 33 equiv) was added. After the reaction had been stirring for 1 h, the volatiles were removed. The crude mixture was dissolved in dry DMF (1 mL) and sulfo-Cyanine5 carboxylic acid³⁵ (15 mg, 23 μ mol, 1.0 equiv), EDC (4.6 mg, 30 μ mol, 1.3 equiv), HOBt (4.5 mg, 30 μ mol, 1.3 equiv), DIPEA (8.2 μ L, 46 μ mol, 2.0 equiv), and molecular sieves (4 Å) were added. The reaction was stirred for 16 h, whereupon it was filtered and the volatiles were removed. The crude product was purified using semipreparative HPLC with a $\text{H}_2\text{O}/\text{MeCN}$ gradient containing 1% trifluoroacetic acid (10% MeCN to 60% in 20 min, to 100% in 1 min. R_t = 14.98 min), yielding tetrazine-Cy5 **4** (4.4 mg, 21%) as a blue solid. ^1H NMR (500 MHz, CD_3OD) δ 9.08 (d, J = 2.4 Hz, 1H), 8.89–8.83 (m, 1H), 8.75 (d, J = 8.6 Hz, 1H), 8.70 (dt, J = 7.9, 1.1 Hz, 1H), 8.49 (dd, J = 8.7, 2.6 Hz, 1H), 8.33–8.28 (m, 2H), 8.16 (td, J = 7.8, 1.7 Hz, 1H), 7.93–7.86 (m, 4H), 7.72 (ddd, J = 7.7, 4.8, 1.2 Hz,

1H), 7.37 (d, J = 8.2 Hz, 1H), 7.30 (d, J = 8.3 Hz, 1H), 6.65 (t, J = 12.3 Hz, 1H), 6.39–6.35 (m, 1H), 6.29 (d, J = 13.7 Hz, 1H), 4.20–4.13 (m, 4H), 4.05 (s, 2H), 2.39–2.31 (m, 2H), 1.92–1.84 (m, 2H), 1.79–1.73 (m, 14H), 1.58–1.49 (m, 2H), 1.42–1.37 (m, 2H). ^{13}C NMR (125 MHz, CD_3OD) δ 175.1, 174.0, 173.6, 169.1, 163.3, 163.1, 154.9, 154.8, 150.1, 149.8, 144.1, 143.6, 143.1, 142.04, 141.96, 141.34, 141.32, 141.2, 138.7, 138.2, 127.1, 126.8, 126.7, 126.2, 124.8, 124.2, 120.0, 119.9, 110.4, 110.0, 103.9, 103.5, 49.21, 49.20, 43.7, 42.6, 38.8, 34.8, 26.6, 26.4, 26.30, 26.29, 25.6, 24.8, 11.1. HRMS (ESI+) m/z calcd. for $\text{C}_{47}\text{H}_{50}\text{N}_{10}\text{O}_8\text{S}_2$ [$\text{M} + \text{H}$]⁺ 947.33327, found: 947.33451.

Expression of ELP-CCMV. The pET-15b-G-H₆-[V₄L₄G₁-9]-CCMV(Δ N26) vector encoding for the hexahistidine-tagged ELP-CCMV protein was previously constructed as described by van Eldijk et al.³ The expression was performed according to a literature procedure.¹⁵ For a typical expression, LB medium (50 mL), containing ampicillin (100 mg/L) and chloramphenicol (50 mg/L), was inoculated with a single colony of *E. coli* BLR(DE3)pLysS containing the pET-15b vector encoding for the ELP-CCMV capsid protein, and was incubated overnight at 37 °C. This overnight culture was used to inoculate 2 \times TY medium (1 L), supplemented with ampicillin (100 mg/L). The culture was grown at 37 °C and protein expression was induced during logarithmic growth (OD_{600} = 0.4–0.6) by addition of IPTG (1 mM). After 6 h of expression at 30 °C, the cells were harvested by centrifugation (2700 g, 15 min, 4 °C) and the pellets were stored overnight at –20 °C.

After thawing, the cell pellet was resuspended in lysis buffer (50 mM NaH_2PO_4 , 1.3 M NaCl, 10 mM imidazole, pH 8.0; 25 mL). The cells were lysed by ultrasonic disruption (3 times 30 s, 100% duty cycle, output control 3, Branson Sonifier 250, Marius Instruments). Then, the lysate was centrifuged (16 400 g, 15 min, 4 °C) to remove the cellular debris. The supernatant was incubated with Ni-NTA agarose beads (3 mL) for 1 h at 4 °C. The suspension was loaded onto a column, the flow-through was collected and the beads were washed twice with wash buffer (50 mM NaH_2PO_4 , 1.3 M NaCl, 20 mM imidazole, pH 8.0; 20 mL). Then, the protein of interest was eluted from the column with elution buffer (50 mM NaH_2PO_4 , 1.3 M NaCl, 250 mM imidazole, pH 8.0; 1 time 0.5 mL, 7 times 1.5 mL). The purification was analyzed by SDS-PAGE. The fractions containing the desired protein were combined and dialyzed against pH 7.5 dimer buffer to obtain the capsid protein dimers. For storage, the proteins were assembled by dialysis against pH 5.0 capsid buffer. The pure protein was obtained with a yield of 100 mg/L of bacterial culture. The purity of the proteins was verified by SDS-PAGE. The assembly properties of the capsid proteins and the geometry of the resulting capsids were analyzed by SEC using a Superose 6 GL 10/300 column with pH 5.0 capsid buffer as the eluent and by TEM. ESI-TOF: calculated 22 253.4 Da, found 22 253.5 Da.

Stability Studies of ELP-CCMV. ELP-CCMV was dialyzed to PBS buffer by spin filtration (10 kDa MWCO, 3 \times 10 min) and diluted to 50 and 10 μ M. Next, the samples (25 μ L) were incubated at 21, 25, 30, and 37 °C for 24 h (400 rpm), whereupon the samples were centrifuged (1 min, 13 000 rpm) and the supernatant was transferred to a clean Eppendorf tube. Loss of protein in the form of precipitation/aggregation was monitored by loading the soluble protein fraction onto an SDS-PAGE gel, the 50 μ M samples were diluted 5 \times to be able to compare them to the 10 μ M samples (Figure S3).

Protein Modification with 2PCA 2. For a typical modification using 2PCA 2, a stock solution of ELP-CCMV in PBS buffer was prepared by spin filtration to this buffer (10 kDa MWCO, 3 × 10 min). The protein (50 μM) and the indicated concentration of 2PCA 2 (100× stock in DMSO, 0 to 100 equiv) were combined in PBS buffer and incubated at 21 °C for 24 h (400 rpm). The samples were analyzed by ESI-TOF (Figure 3B, C, D and Figure S4, S5) and SDS-PAGE (Figure 3E).

Protein Modification with 2PCA 2 or 2PCA-VBA 3, Followed by Tetrazine Ligation. ELP-CCMV was dialyzed to PBS by spin filtration (10 kDa MWCO, 3 × 10 min) and diluted to 10 μM. Then, 2PCA 2 or 2PCA-VBA 3 (100 mM, 100× in DMSO, 100 equiv) or DMSO were added to the protein (10 μM) and the samples were incubated at 21 °C for 24 h (400 rpm). The samples were centrifuged (1 min, 13 000 rpm), after which they were dialyzed with PBS buffer to remove the excess of the small molecule (Spectra/Por 4 dialysis tubing, 12–14 kDa MWCO, 10 mm flat width, 3 × 60 min). Next, tetrazine-Cy5 4 (100 μM, 10 equiv, 10 mM stock solution in DMSO) or DMSO was added to the protein (10 μM) and the samples were incubated at 21 °C for 1 h. The protein modification steps were analyzed by SDS-PAGE (Figure 4C), whereas the protein modification step using 2PCA-VBA 3 (100 equiv) was also analyzed by ESI-TOF (Figure S6).

Protein Modification with a Concentration Range of 2PCA-VBA 3, Followed by Tetrazine Ligation. The concentration range was performed using the same method as described for “Protein modification with 2PCA 2 or 2PCA-VBA 3, followed by tetrazine ligation” only the indicated concentration of 2PCA-VBA 3 (100× solution in DMSO, 0 to 250 equiv) was added to ELP-CCMV (10 μM) in the first step of the modification. The samples were then analyzed by SDS-PAGE (Figure 4D).

■ ASSOCIATED CONTENT

■ Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.bioconjchem.7b00815.

Experimental details for the synthesis and expression, modification and capsid formation of CCMV, full spectroscopic data for all new compounds and additional figures (PDF)

■ AUTHOR INFORMATION

Corresponding Authors

*E-mail: k.bonger@science.ru.nl.

*E-mail: j.c.m.v.hest@tue.nl.

ORCID

Kimberly M. Bonger: 0000-0001-9498-2620

Jan C. M. van Hest: 0000-0001-7973-2404

Author Contributions

#Lise Schoonen and Selma Eising contributed equally to this work.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

Mr. Floris van Dalen and Dr. Martijn Verdoes are kindly acknowledged for the synthesis and characterization of sulfo-

Cyanine5 carboxylic acid. L. Schoonen, M. B. van Eldijk, R. J. M. Nolte, and J. C. M. van Hest thank the Ministry of Education, Culture and Science (NWO Gravitation program 024.001.035) for financial support. R. J. M. Nolte also thanks the European Research Council (ERC Advanced Grant, 740295 ENCOPOL). S. Eising and K. M. Bonger acknowledge funding from The Netherlands Research Institute for Chemical Biology (NRSCB) and the Institute of Molecules and Materials (IMM) of the Radboud University in Nijmegen.

■ REFERENCES

- (1) Caspar, D. L. D., and Klug, A. (1962) Physical Principles in the Construction of Regular Viruses. *Cold Spring Harbor Symp. Quant. Biol.* 27, 1–24.
- (2) Speir, J. A., Munshi, S., Wang, G., Baker, T. S., and Johnson, J. E. (1995) Structures of the native and swollen forms of cowpea chlorotic mottle virus determined by X-ray crystallography and cryo-electron microscopy. *Structure* 3, 63–78.
- (3) van Eldijk, M. B., Wang, J. C.-Y., Minten, I. J., Li, C., Zlotnick, A., Nolte, R. J. M., Cornelissen, J. J. L. M., and van Hest, J. C. M. (2012) Designing Two Self-Assembly Mechanisms into One Viral Capsid Protein. *J. Am. Chem. Soc.* 134, 18506–18509.
- (4) Urry, D. W. (1997) Physical Chemistry of Biological Free Energy Transduction As Demonstrated by Elastic Protein-Based Polymers. *J. Phys. Chem. B* 101, 11007–11028.
- (5) Urry, D. W. (2004) The change in Gibbs free energy for hydrophobic association: Derivation and evaluation by means of inverse temperature transitions. *Chem. Phys. Lett.* 399, 177–183.
- (6) Comellas-Aragonès, M., Engelkamp, H., Claessen, V. I., Sommerdijk, N. A. J. M., Rowan, A. E., Christianen, P. C. M., Maan, J. C., Verduin, B. J. M., Cornelissen, J. J. L. M., and Nolte, R. J. M. (2007) A virus-based single-enzyme nanoreactor. *Nat. Nanotechnol.* 2, 635–639.
- (7) Cadena-Nava, R. D., Hu, Y., Garmann, R. F., Ng, B., Zelikin, A. N., Knobler, C. M., and Gelbart, W. M. (2011) Exploiting fluorescent polymers to probe the self-assembly of virus-like particles. *J. Phys. Chem. B* 115, 2386–2391.
- (8) Setaro, F., Brasch, M., Hahn, U., Koay, M. S. T., Cornelissen, J. J. L. M., De La Escosura, A., and Torres, T. (2015) Generation-Dependent Templated Self-Assembly of Biohybrid Protein Nanoparticles around Photosensitizer Dendrimers. *Nano Lett.* 15, 1245–1251.
- (9) Brasch, M., Putri, R. M., de Ruiter, M. V., Luque, D., Koay, M. S. T., Castón, J. R., and Cornelissen, J. J. L. M. (2017) Assembling enzymatic cascade pathways inside virus-based nanocages using dual-tasking nucleic acid tags. *J. Am. Chem. Soc.* 139, 1512–1519.
- (10) Rurup, W. F., Verbij, F., Koay, M. S. T., Blum, C., Subramaniam, V., and Cornelissen, J. J. L. M. (2014) Predicting the loading of virus-like particles with fluorescent proteins. *Biomacromolecules* 15, 558–563.
- (11) Minten, I. J., Hendriks, L. J. A., Nolte, R. J. M., and Cornelissen, J. J. L. M. (2009) Controlled encapsulation of multiple proteins in virus capsids. *J. Am. Chem. Soc.* 131, 17771–17773.
- (12) Minten, I. J., Claessen, V. I., Blank, K., Rowan, A. E., Nolte, R. J. M., and Cornelissen, J. J. L. M. (2011) Catalytic capsids: the art of confinement. *Chem. Sci.* 2, 358–362.
- (13) Schoonen, L., Pille, J., Borrmann, A., Nolte, R. J. M. R. J. M., and van Hest, J. C. M. J. C. M. (2015) Sortase A-mediated N-terminal modification of cowpea chlorotic mottle virus for highly efficient cargo loading. *Bioconjugate Chem.* 26, 2429–2434.
- (14) Schoonen, L., Nolte, R. J. M., and van Hest, J. C. M. (2016) Highly efficient enzyme encapsulation in a protein nanocage: towards enzyme catalysis in a cellular nanocompartment mimic. *Nanoscale* 8, 14467–14472.
- (15) Sletten, E. M., and Bertozzi, C. R. (2009) Bioorthogonal chemistry: Fishing for selectivity in a sea of functionality. *Angew. Chem., Int. Ed.* 48, 6974–6998.

- (16) Gong, Y., and Pan, L. (2015) Recent advances in bioorthogonal reactions for site-specific protein labeling and engineering. *Tetrahedron Lett.* 56, 2123–2132.
- (17) Krall, N., da Cruz, F. P., Boutureira, O., and Bernardes, G. J. L. (2016) Site-selective protein-modification chemistry for basic biology and drug development. *Nat. Chem.* 8, 103–113.
- (18) Rosen, C. B., and Francis, M. B. (2017) Targeting the N terminus for site-selective protein modification. *Nat. Chem. Biol.* 13, 697–705.
- (19) Kiick, K. L., Saxon, E., Tirrell, D. A., and Bertozzi, C. R. (2002) Incorporation of azides into recombinant proteins for chemoselective modification by the Staudinger ligation. *Proc. Natl. Acad. Sci. U. S. A.* 99, 19–24.
- (20) Schoffelen, S., Lambermon, M. H. L., van Eldijk, M. B., and van Hest, J. C. M. (2008) Site-specific modification of *Candida antarctica* lipase B via residue-specific incorporation of a non-canonical amino acid. *Bioconjugate Chem.* 19, 1127–1131.
- (21) Chin, J. W., Santoro, S. W., Martin, A. B., King, D. S., Wang, L., and Schultz, P. G. (2002) Addition of p-azido-L-phenylalanine to the genetic code of *Escherichia coli*. *J. Am. Chem. Soc.* 124, 9026–9027.
- (22) Tanrikulu, I. C., Schmitt, E., Mechulam, Y., Goddard, W. A., and Tirrell, D. A. (2009) Discovery of *Escherichia coli* methionyl-tRNA synthetase mutants for efficient labeling of proteins with azidonorleucine in vivo. *Proc. Natl. Acad. Sci. U. S. A.* 106, 15285–15290.
- (23) van Dongen, S. F. M., Teeuwen, R. L. M., Nallani, M., van Berkel, S. S., Cornelissen, J. J. L. M., Nolte, R. J. M., and van Hest, J. C. M. (2009) Single-Step Azide Introduction in Proteins via an Aqueous Diazo Transfer. *Bioconjugate Chem.* 20, 20–23.
- (24) Schoffelen, S., van Eldijk, M. B., Rooijakkers, B., Raijmakers, R., Heck, A. J. R., and van Hest, J. C. M. (2011) Metal-free and pH-controlled introduction of azides in proteins. *Chem. Sci.* 2, 701–705.
- (25) Kulkarni, C., Kinzer-ursem, T. L., Tirrell, D. A., Gmbh, C. W. V., Kgaa, C., Kulkarni, C., Kinzer-ursem, T. L., and Tirrell, D. A. (2013) Selective Functionalization of the Protein N Terminus with N-Myristoyl Transferase for Bioconjugation in Cell Lysate. *ChemBioChem* 14, 1958–1962.
- (26) Laughlin, S. T., and Bertozzi, C. R. (2007) Metabolic labeling of glycans with azido sugars and subsequent glycan-profiling and visualization via Staudinger ligation. *Nat. Protoc.* 2, 2930–2944.
- (27) Debets, M. F., van Hest, J. C. M., and Rutjes, F. P. J. T. (2013) Bioorthogonal labelling of biomolecules: new functional handles and ligation methods. *Org. Biomol. Chem.* 11, 6439–6455.
- (28) Saxon, E. (2000) Cell Surface Engineering by a Modified Staudinger Reaction. *Science* 287, 2007–2010.
- (29) Dalboge, H., Bayne, S., and Pedersen, J. (1990) In vivo processing of N-terminal methionine in *E. coli*. *FEBS Lett.* 266, 1–3.
- (30) Debets, M. F., van Berkel, S. S., Dommerholt, J., Dirks, A. J., Rutjes, F. P. J. T., and van Delft, F. L. (2011) Bioconjugation with strained alkenes and alkynes. *Acc. Chem. Res.* 44, 805–815.
- (31) MacDonald, J. I., Munch, H. K., Moore, T., and Francis, M. B. (2015) One-step site-specific modification of native proteins with 2-pyridinecarboxyaldehydes. *Nat. Chem. Biol.* 11, 326–331.
- (32) Knall, A.-C., and Slugovc, C. (2013) Inverse electron demand Diels–Alder (iEDDA)-initiated conjugation: a (high) potential click chemistry scheme. *Chem. Soc. Rev.* 42, 5131–5142.
- (33) Oliveira, B. L., Guo, Z., and Bernardes, G. J. L. (2017) Inverse electron demand Diels–Alder reactions in chemical biology. *Chem. Soc. Rev.* 46, 4895–4950.
- (34) Eising, S., Lelivelt, F., and Bongers, K. M. (2016) Vinylboronic Acids as Fast Reacting, Synthetically Accessible, and Stable Bioorthogonal Reactants in the Carboni-Lindsey Reaction. *Angew. Chem., Int. Ed.* 55, 12243–12247.
- (35) Mujumdar, R. B., Ernst, L. A., Mujumdar, S. R., Lewis, C. J., and Waggoner, A. S. (1993) Cyanine Dye Labeling Reagents: Sulfoindocyanine Succinimidyl Esters. *Bioconjugate Chem.* 4, 105–111.